

# Package: SIEVEseq (via r-universe)

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**Title** Unified Differential Expression, Variability and Skewness  
Analysis for RNA-Seq Data

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**Description** Provides a unified framework for the simultaneous testing of differential expression, variability, and skewness of genes using RNA-Seq data. The framework adopts a compositional data analysis approach for modelling RNA-Seq count data, applies the centered log-ratio transformation to obtain continuous variables, and uses the skew-normal distribution for statistical inference. Methods are described in Li and Khang (bioRxiv preprint, 2024, version 3) <doi:10.1101/2024.04.09.588804>.

**Depends** R (>= 4.1)

**Imports** sn, stats, ggplot2

**Suggests** knitr, rmarkdown, testthat (>= 3.0.0)

**VignetteBuilder** knitr

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**License** GPL (>= 2)

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clr.SN.fit	<i>Fit the skew-normal distribution to CLR-transformed RNA-Seq data</i>
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### Description

Estimate the mean, standard deviation, and skewness parameters of the skew-normal distribution using centered log-ratio (CLR) transformed RNA-Seq data.

### Usage

```
clr.SN.fit(data)
```

### Arguments

data	A table of CLR-transformed count data, with genes/transcripts on the rows and samples on columns.
------	---

### Value

mu	The maximum likelihood estimate of the mean parameter.
se.mu	The standard error of the maximum likelihood estimate of mu.
z.mu	The Wald statistic for mu.
p.mu	The p-value of the Wald statistic for mu.
sigma	The maximum likelihood estimate of the standard deviation parameter.
se.sigma	The standard error of the maximum likelihood estimate of sigma.
z.sigma	The Wald statistic for sigma.
p.sigma	The p-value of the Wald statistic for sigma.
gamma	The maximum likelihood estimate of the skewness parameter.
se.gamma	The standard error of the maximum likelihood estimate of gamma.
z.gamma	The Wald statistic for gamma.
p.gamma	The p-value of the Wald statistic for gamma.

## References

Azzalini, A. (1985). *A class of distributions which includes the normal ones*. *Scandinavian Journal of Statistics* **12**(2), 171–178, JSTOR

Azzalini, A. and Capitanio, A. (2014). *The Skew-Normal and Related Families*. Cambridge University Press, IMS Monographs series.

Azzalini, A. and Arellano-Valle, R. B. (2013). Maximum penalized likelihood estimation for skew-normal and skew-*t* distributions. *Journal of Statistical Planning and Inference* **143**, 419–433.

Azzalini, A. (2022). *The R package sn: The skew-normal and related distribution such as the skew-t and the SUN (version 2.0.2)*. Università degli Studi di Padova, Italia. Home page: <https://cran.r-project.org/package=sn>.

Aitchison, J. (1986). *The Statistical Analysis of Compositional Data*. Chapman & Hall, London.

## Examples

```
library(SIEVEseq)
data(clrCounts1)
clr.SN.fit(clrCounts1[1:2, ])
clr.SN.fit(clrCounts1[1, ])
```

---

clrCounts1

*Simulated CLR-transformed count table*

---

## Description

A simulated CLR-transformed count table with genes on the rows and samples on the columns for one condition.

## Usage

```
clrCounts1
```

## Format

A data frame containing CLR-transformed counts, with 200 genes and 500 samples.

---

clrCounts2	<i>Simulated DV genes CLR-transformed count table</i>
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---

**Description**

A simulated CLR-transformed count table with genes on the rows and samples on the columns for two groups. Ten percent of the genes are DV genes (gene1 to gene50).

**Usage**

```
clrCounts2
```

**Format**

A data frame containing CLR-transformed counts, with 500 genes and 200 samples per group.

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clrCounts3	<i>Simulated DE genes CLR-transformed count table</i>
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---

**Description**

A simulated CLR-transformed count table with genes on the rows and samples on the columns for two groups. Ten percent of the genes are DE genes (gene1 to gene50).

**Usage**

```
clrCounts3
```

**Format**

A data frame containing CLR-transformed counts, with 500 genes and 200 samples per group.

---

`clrDE`*Differential Expression (DE) Test of CLR-transformed RNA-Seq Data*

---

**Description**

Model CLR-transformed RNA-Seq data using the skew-normal distribution, and then conduct a statistical test for finding genes/transcripts with differential expression using the Wald test.

**Usage**

```
clrDE(data = NULL, group = NULL)
```

**Arguments**

<code>data</code>	CLR-transformed counts matrix with genes/transcripts on the rows and samples on the columns
<code>group</code>	2 groups (control vs. treatment)

**Value**

An object of 'clrDE' class that contains the results of the DV test and associated information:

<code>DE</code>	The difference of mu between group 2 and group 1 ( $\mu_2 - \mu_1$ ).
<code>se</code>	The standard error of DE.
<code>z</code>	The observed Wald statistic.
<code>pval</code>	The unadjusted p-value of the Wald test.
<code>adj_pval</code>	The p-value of the Wald test adjusted using the Benjamini-Yekutieli procedure.
<code>mu1</code>	The maximum likelihood estimate of the mean parameter of group 1.
<code>se.mu1</code>	The standard error of the maximum likelihood estimate of $\mu_1$ .
<code>z.mu1</code>	The Wald statistic for $\mu_1$ .
<code>p.mu1</code>	The p-value of the Wald test for $\mu_1$ .
<code>mu2</code>	The maximum likelihood estimate of the mean parameter of group 2.
<code>se.mu2</code>	The standard error of the maximum likelihood estimate of $\mu_2$ .
<code>z.mu2</code>	The Wald statistic for $\mu_2$ .
<code>p.mu2</code>	The p-value of the Wald test for $\mu_2$ .

**See Also**

[clrSIEVE\(\)](#) for DE, DV and DS tests.

**Examples**

```

library(SIEVEseq)
data(clrCounts3) # The first 50 genes (gene1 to gene50) are DE genes
groups <- c(rep(0, 200), rep(1, 200))
clrDE_test <- clrDE(clrCounts3[46:100, ], group = groups)
sum(is.na(clrDE_test)) # check NA values
head(clrDE_test, 5) # adj_pval < 0.05, DE genes
tail(clrDE_test, 5) # adj_pval > 0.05, non-DE genes

```

---

 clrDV

*Differential variability (DV) test of CLR-transformed RNA-Seq data*


---

**Description**

Model CLR-transformed RNA-Seq data using the skew-normal distribution, and then conduct a statistical test for finding genes/transcripts with differential variability using the Wald test.

**Usage**

```
clrDV(data = NULL, group = NULL)
```

**Arguments**

data	A CLR-transformed count matrix with genes/transcripts on the rows and samples on the columns.
group	A vector specifying the group labels of the data.

**Value**

An object of 'clrDV' class that contains the results of the DV test and associated information:

DV	The difference of standard deviation ( $\sigma$ ) between group 2 and group 1 ( $\sigma_2 - \sigma_1$ ).
se	The standard error of DV.
z	The observed Wald statistic.
pval	The unadjusted p-value of Wald test.
adj_pval	The p-value of the Wald test adjusted using the Benjamini-Yekutieli procedure.
sigma1	The maximum likelihood estimate of the standard deviation parameter for group 1.
se.sigma1	The standard error of the maximum likelihood estimate of $\sigma_1$ .
z.sigma1	The Wald statistic for $\sigma_1$ .
p.sigma1	The p-value of the Wald test for $\sigma_1$ .
sigma2	The maximum likelihood estimate of the standard deviation parameter for group 2.

se.sigma2	The standard error of the maximum likelihood estimate of gamma2.
z.sigma2	The Wald statistic for sigma2.
p.sigma2	The p-value of the Wald test for sigma2.

**See Also**

[clrSIEVE\(\)](#) for DE, DV and DS tests.

**Examples**

```
library(SIEVEseq)
data(clrCounts2) # first 50 genes (gene1 to gene50) are DV genes
groups <- c(rep(0, 200), rep(1, 200))
clrDV_test <- clrDV(clrCounts2[46:100, ], group = groups)
sum(is.na(clrDV_test)) # check NA values
head(clrDV_test, 5) # adj_pval < 0.05, DV genes
tail(clrDV_test, 5) # adj_pval > 0.05, non-DV genes
```

---

clrSeq	<i>Fit the skew-normal model to CLR-transformed RNA-Seq data for 2 groups</i>
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---

**Description**

Estimate the mean, standard deviation, and skewness parameters of the skew-normal distribution using centered log-ratio (CLR) transformed RNA-Seq data for 2 groups. The related computational works (standard error, Wald statistic, and p value) are also provided.

**Usage**

```
clrSeq(data = NULL, group = NULL)
```

**Arguments**

data	A table of CLR-transformed count data, with genes/transcripts on the rows and samples on the columns for 2 groups.
group	A vector specifying the group labels of the data.

**Value**

mu1	The maximum likelihood estimate of the mean parameter of group 1.
se.mu1	The standard error of the maximum likelihood estimate of mu1.
z.mu1	The Wald statistic for mu1.
p.mu1	The p-value of the Wald test for mu1.

sigma1	The maximum likelihood estimate of the standard deviation parameter of group 1.
se.sigma1	The standard error of the maximum likelihood estimate of sigma1.
z.sigma1	The Wald statistic for sigma1.
p.sigma1	The p-value of the Wald test for sigma1.
gamma1	The maximum likelihood estimate of the skewness parameter of group 1.
se.gamma1	The standard error of the maximum likelihood estimate of gamma1.
z.gamma1	The Wald statistic for gamma1.
p.gamma1	The p-value of the Wald test for gamma1.
mu2	The maximum likelihood estimate of mean parameter of group 2.
se.mu2	The standard error of the maximum likelihood estimate of mu2.
z.mu2	The Wald statistic for mu2.
p.mu2	The p-value of the Wald test for mu2.
sigma2	The maximum likelihood estimate of the standard deviation parameter of group 2.
se.sigma2	The standard error of the maximum likelihood estimate of sigma2.
z.sigma2	The Wald statistic for sigma2.
p.sigma2	The p-value of the Wald test for sigma2.
gamma2	The maximum likelihood estimate of the skewness parameter of group 2.
se.gamma2	The standard error of the maximum likelihood estimate of gamma2.
z.gamma2	The Wald statistic for gamma2.
p.gamma2	The p-value of the Wald test for gamma2.

## References

- Azzalini, A. (1985). *A class of distributions which includes the normal ones*. *Scandinavian Journal of Statistics* **12**(2), 171–178, JSTOR
- Azzalini, A. and Capitanio, A. (2014). *The Skew-Normal and Related Families*. Cambridge University Press, IMS Monographs series.
- Azzalini, A. and Arellano-Valle, R. B. (2013). Maximum penalized likelihood estimation for skew-normal and skew-*t* distributions. *Journal of Statistical Planning and Inference* **143**, 419–433.
- Azzalini, A. (2022). *The R package sn: The skew-normal and related distribution such as the skew-t and the SUN (version 2.0.2)*. Università degli Studi di Padova, Italia. Home page: <https://cran.r-project.org/package=sn>.
- Aitchison, J. (1986). *The Statistical Analysis of Compositional Data*. Chapman & Hall, London.

## See Also

`clrSIEVE()` for DE, DV and DS tests.

**Examples**

```

library(SIEVEseq)
data("clrCounts2")
groups <- c(rep(0, 200), rep(1, 200))
clrSeq_result <- clrSeq(clrCounts2[46:100, ], group = groups)
tail(clrSeq_result, 5)
SN.plot(clrCounts2[1, 1:200])
clrSeq_result[1, c("mu1", "sigma1", "gamma1")]

```

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clrSIEVE	<i>Simultaneous Differential Expression, Variability and Skewness Analysis Using RNA-Seq Data</i>
----------	---

---

**Description**

Model CLR-transformed RNA-Seq data using the skew-normal distribution, and then conduct statistical tests for finding genes/transcripts with differential expression, variability and skewness using the Wald test.

**Usage**

```

clrSIEVE(
  clrSeq_result = NULL,
  alpha_level = 0.05,
  order_DE = FALSE,
  order_LFC = FALSE,
  order_DS = FALSE,
  order_sieve = FALSE
)

```

**Arguments**

clrSeq_result	The result of clrSeq() function.
alpha_level	The adjusted p-value cutoff used for flagging genes that show significant differential expression, variability and skewness.
order_DE	Logical string. "FALSE" for no ordering; "TRUE" for ordering by the value of DE.
order_LFC	Logical string. "FALSE" for no ordering; "TRUE" for ordering by the value of LFC.
order_DS	Logical string. "FALSE" for no ordering; "TRUE" for ordering by the value of DS.
order_sieve	Character/logical string specifying the order method. Possibilities are "DE" for the value of DE, "LFC" for log <sub>2</sub> fold change of variability, "DS" for the value of DS or "FALSE" for no ordering.

**Value**

clrDE\_test, clrDV\_test and clrDS\_test contain the result of the DE, DV and DS tests, respectively. clrSIEVE\_tests contains the result of all three tests.

clrDE_test	A data.frame contating the results of differential expression test.
clrDV_test	A data.frame contating the results of differential variability test.
clrDS_test	A data.frame contating the results of differential skewness test.
clrSIEVE_tests	A data.frame contating the results of differential expression, variability and skewness tests.
DE	The difference of mean ( $\mu$ ) between group 2 and group 1 ( $\mu_2 - \mu_1$ ).
se_DE	The standard error of DE.
z_DE	The observed Wald statistic of DE.
pval_DE	The unadjusted p-value of Wald test of DE.
adj_pval_DE	The p-value of the Wald test of DE, adjusted using the Benjamini-Yekutieli procedure.
mu1	The maximum likelihood estimate of the mean parameter for group 1.
mu2	The maximum likelihood estimate of the mean parameter for group 2.
de_indicator	1: DE gene; 0: non-DE gene.
SD_ratio	The ratio of standard deviation ( $\sigma$ ) between group 2 and group 1 ( $\sigma_2/\sigma_1$ ).
LFC	log2 fold change: $\log_2(\sigma_2/\sigma_1)$ .
DV	The difference of standard deviation ( $\sigma$ ) between group 2 and group 1 ( $\sigma_2 - \sigma_1$ ).
se_DV	The standard error of DV.
z_DV	The observed Wald statistic of DV.
pval_DV	The unadjusted p-value of Wald test of DV.
adj_pval_DV	The p-value of the Wald test of DV, adjusted using the Benjamini-Yekutieli procedure.
sigma1	The maximum likelihood estimate of the standard deviation parameter for group 1.
sigma2	The maximum likelihood estimate of the standard deviation parameter for group 2.
dv_indicator	1: DV gene; 0: non-DV gene.
DS	The difference of skewness parameter ( $\gamma$ ) between group 2 and group 1 ( $\gamma_2 - \gamma_1$ ).
se_DS	The standard error of DS.
z_DS	The observed Wald statistic of DS.
pval_DS	The unadjusted p-value of Wald test of DS.
adj_pval_DS	The p-value of the Wald test of DS adjusted using the Benjamini-Yekutieli procedure.
gamma1	The maxiimum likelihood estimate of the skewness parameter for group 1.
gamma2	The maxiimum likelihood estimate of the skewness parameter for group 2.
ds_indicator	1: DS gene; 0: non-DS gene.

## References

- Azzalini, A. (1985). *A class of distributions which includes the normal ones*. *Scandinavian Journal of Statistics* **12**(2), 171–178, JSTOR
- Azzalini, A. and Capitanio, A. (2014). *The Skew-Normal and Related Families*. Cambridge University Press, IMS Monographs series.
- Azzalini, A. and Arellano-Valle, R. B. (2013). Maximum penalized likelihood estimation for skew-normal and skew-*t* distributions. *Journal of Statistical Planning and Inference* **143**, 419–433.
- Azzalini, A. (2022). *The R package sn: The skew-normal and related distribution such as the skew-t and the SUN (version 2.0.2)*. Università degli Studi di Padova, Italia. Home page: <https://cran.r-project.org/package=sn>.
- Aitchison, J. (1986). *The Statistical Analysis of Compositional Data*. Chapman & Hall, London.

## See Also

`clrSeq()` for the results of parameters estimation; alternatively, `clrDE()` only provides DE test and `clrDV()` only provides DV test.

## Examples

```
library(SIEVEseq)
data(clrCounts3) # first 50 genes (gene1 to gene50) are DE genes
groups <- c(rep(0, 200), rep(1, 200))
clrSeq_result3 <- clrSeq(clrCounts3[46:100, ], group = groups) # DE dataset
clrSIEVE_result3 <- clrSIEVE(clrSeq_result = clrSeq_result3,
                             alpha_level = 0.05,
                             order_DE = FALSE,
                             order_LFC = FALSE,
                             order_DS = FALSE,
                             order_sieve = FALSE)
clrDE_test3 <- clrSIEVE_result3$clrDE_test # DE test
head(clrDE_test3, 5)
clrDS_test3 <- clrSIEVE_result3$clrDS_test # DS test
clrDS_test3[clrDS_test3$adj_pval_DS < 0.05, ]
clrSIEVE_tests3 <- clrSIEVE_result3$clrSIEVE_tests # Sieve DE, DV and DS genes
head(clrSIEVE_tests3, 5)
tail(clrSIEVE_tests3, 5)
```

## Description

Produce a histogram of observed CLR-transformed counts, with the fitted skew-normal probability density function for a particular gene/transcript.

**Usage**

```
SN.plot(data)
```

**Arguments**

data            CLR-transformed counts for a particular gene/transcript.

**Value**

No return value. This function is called for its side effect of generating a plot showing the observed CLR-transformed counts and the fitted skew-normal density.

**Examples**

```
library(SIEVEseq)
data("clrCounts1")
SN.plot(clrCounts1[1,])
SN.plot(clrCounts1[2,])
SN.plot(clrCounts1[3,])
```

---

violin.plot.SIEVE      *Violin plots*

---

**Description**

Produce violin plots of CLR-transformed count data for two or three groups.

**Usage**

```
violin.plot.SIEVE(  
  data = NULL,  
  name.gene = NULL,  
  group = NULL,  
  group.names = NULL,  
  xlab = "CLR-transformed count",  
  ylab = "Condition"  
)
```

**Arguments**

data            A CLR-transformed count table.  
name.gene       Gene/transcript name.  
group           A vector specifying the group labels of the data.  
group.names     A vector specifying the group names.  
xlab            Name of the x-axis.  
ylab            Name of the y-axis.

**Value**

A ggplot object displaying the distribution of CLR-transformed expression values for the specified gene across groups.

**Examples**

```
library(SIEVEseq)
data(clrCounts2) # first 50 genes (gene1 to gene50) are DV genes
data(clrCounts3) # first 50 genes (gene1 to gene50) are DE genes
group0 <- c(rep(0, 200), rep(1, 200))
group1 <- c(rep(0, 200), rep(1, 100), rep(2, 100))
violin.plot.SIEVE(data = clrCounts2, "gene1", group = group0,
  group.names = c("control", "case")) # DV
violin.plot.SIEVE(data = clrCounts2, "gene1", group = group1,
  group.names = c("control", "case1", "case2")) # DV
violin.plot.SIEVE(data = clrCounts3, "gene1", group = group0,
  group.names = c("control", "case")) # DE
violin.plot.SIEVE(data = clrCounts3, "gene2", group = group0,
  group.names = c("control", "case")) # DE
violin.plot.SIEVE(data = clrCounts3, "gene200", group = group0,
  group.names = c("control", "case")) # non-DE
```

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